

ARTICLE / INVESTIGACIÓN

Isolation and identification of intestinal parasites from Goats in some areas of Wasit Province, Iraq

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Abstract: The current study was conducted on goats in various parts of Wasit Province, Iraq, from November 2021 to April 2022. The study aims to find and identify intestinal parasites (IPs) in goats in Wasit province. The goat's fresh fecal specimens (n=180) include cysts, eggs, oocysts, trophozoites and larval stages. One hundred eighty sheep feces samples were collected, and more than one parasite was isolated from one sample (mixed infection). According to the data acquired, the overall prevalence of intestinal parasites in goats was 52.77 (95 samples). In the current investigation, eleven distinct (IPs) species with infection rates were identified, including *Toxocara vitulorum* (Goeze, 1782) (16.66%), *Cryptosporidium* sp. (Tyzzer, 1907) (11.11%), *Amoeba* sp. (8.8%), *Giardia* sp. (Künstler, 1882) (8.8%), *Trichostrongylus* sp. (Looss, 1905) (8.33%), *Cyclospora* sp. (Schneider, 1881) (5.55%), *Dicrocoelium dendriticum* synonym (*Distoma dendriticum*) (Rudolphi, 1819) (5%), *Paramphistomum cervi* (Zeder, 1790) (4.44%), cercaria larva (2.22%), *Balantidium coli* (Malmsten, 1857) (1.66%), filariae form larvae (1.66%), respectively. This may be the result of infection with various parasites due to the use of Conventional and unsanitary management systems

Key words: Intestinal Parasites, Goats, *Toxocara vitulorum*, *Cryptosporidium* spp.

Introduction

The domestic goat was one of the first animals that humans domesticated. They are found all across the planet, with higher abundance in tropical and dry zones¹. Gastrointestinal parasitism is one of the most severe diseases affecting goats, particularly nematode infections, among the most significant health issues limiting the animal's productivity^{2,3}. However, parasites such as trematode (flake) and cestode (tapeworm) may also contribute to harmful worm burdens in animals⁴. Protozoan infections such as amoebiasis, giardiasis, and coccidiosis have been documented in Thailand, Costa Rica, and India^{5,6}. The distribution of most parasitic diseases has been linked to environmental conditions and vector abundance. Gastrointestinal parasites are ubiquitous in temperate and tropical regions but are more prevalent in warm areas with inadequate sanitation and low living standards⁷. The diversity of parasites affects animal health and causes substantial economic losses to the cattle business. It poses a major health risk and a barrier to small ruminant productivity because of the related morbidity, mortality, treatment costs, and control measures⁸. Small ruminant nematode infections cause low production due to stunted growth, poor weight increase, and poor feed consumption⁹. However, the few available studies on isolating intestinal parasites in goats are uncommon in some areas of Wasit Governorate. As a result, the current study aimed to verify the isolation and identification of infection of goats with intestinal parasites raised in these areas.

Materials and methods

This survey was carried out in Wasit province between

November 2021 till April 2022. Fecal samples (n=180) of goats were collected and preserved in transparent, clean, dry, tight-cover sampling containers. Each container was tagged with the relevant data, such as number, time, and date, and maintained in an ice box before being moved to the Natural History Research and Museum Center at the University of Baghdad. The samples were examined microscopically, and parasite data were recorded and maintained at 4 C° until the laboratory exams were completed within 24 hours. The collected fresh feces specimens were tested individually for identifying intestinal parasites utilizing concentration methodology by employing Sugar/salt solution and sedimentation protocol, as reported by (10, 11). In brief, direct smears for eggs/oocysts/ trophozoites were prepared, as well as a sedimentation protocol for eggs and helminths and a flotation method (Scheathers solution) for the detection of nematode eggs and protozoan oocysts.

Results

The overall prevalence of intestinal parasites in goats was 52.77 % over the study period (95 of 180). In Al-Suwaira, the infection rate was 55 percent, while in Al-Shaihe-miyh, the infection rate was 50 % (Table 1). This shows the eggs, cysts, oocysts, and larvae of intestinal parasites found in this investigation (Table 2) (Figureures 1 to 11). It included the following: *Toxocara vitulorum* (16.66%), *Cryptosporidium* sp. (11.11%), *Amoeba* sp. (8.8%), *Giardia* sp. (8.8%), *Trichostrongylus* sp. (8.33%), *Cyclospora* sp. (5.55%), *Dicrocoelium dendriticum* (*Distoma dendriticum*) (5%), *Paramphistomum cervi* (4.44%), cercaria (2.22%),

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Balantidium coli (Malmsten, 1857) Stein, 1863 (1.66%), filariae form larvae (1.66%), respectively. The infection rate with intestinal parasites, according to months recorded higher infection rate was (70%) in April, while the lower infection rate was (33.33%) in December (Table 3). Analysis of the data based on sex revealed females higher infection rate than males, as recorded in females (54.63%) and males (50.60%) (Table 4). The data on mixed intestinal parasite infections shows that most investigated goats had mixed conditions (Table 5). Generally, the highest double illnesses among (*Toxocara vitulorum* + *Cryptosporidium sp*) with a rate (of 2.22%), while the lowest mix infection was recorded between (*Toxocara vitulorum* + *Giardia sp*) with a rate (of 1.11%).

Discussion

Out of the 180 goat fecal samples were collected (one sample for each animal). From the goats examined, 95 (52.77%) were found positive for one or more parasites from Wasit province. The overall higher prevalence of intestinal parasite infection in the areas studied could be attributed to lower host immunity as a result of malnutrition; grazing of young and adult animals together in poorly drained land provide an ideal environment for the transmission of endoparasite eggs to build up a clinical infestation of the host; this finding is consistent with the findings of many other researchers^{12,13}.

Area	No. of Samples Examined	No. of positive	prevalence (%)
Al-Suwaira	60	33	55%
Al-Aziziyah	60	32	53.33%
Al- Shaihemiyh	60	30	50%
Total	180	95	52.77%

Table 1. The prevalence rate of intestinal parasites in goats in the area.

Intestinal parasite	No. of Samples Examined	Phase	No. of positive	prevalence (%)
<i>Toxocara vitulorum</i>	180	Egg	30	16.66%
<i>Cryptosporidium sp</i>		Oocyst	20	11.11%
<i>Amoeba sp</i>		Cyst	16	8.8%
<i>Giardia sp</i>		Cyst	16	8.8%
<i>Trichostrongylus sp</i>		Egg	15	8.33%
<i>Cyclospora sp</i>		Oocysts	10	5.55%
<i>Dicrocoelium dendriticum</i> (<i>Distoma dendriticum</i>)		Egg	9	5%
<i>Paramphistomum cervi</i>		Egg	8	4.44%
Cercaria		larvae	4	2.22%
<i>Balantidium coli</i>		Cyst	3	1.66%
Filariae		larvae	3	1.66%

Table 2. The prevalence rate with intestinal parasites isolated from 180 fecal samples of goats.

Months/2021	No. of Samples Examined	No. of positive	prevalence (%)
November	30	16	53.33%
December	30	10	33.33%
January	30	15	50%
February	30	15	50%
March	30	18	60%
April	30	21	70%
Total	180	95	52.77%

Table 3. The prevalence rate with intestinal parasites according to months of study.

Sex	No. of Samples Examined	No. of positive	prevalence (%)
Male	83	42	50.60%
Female	97	53	54.63%
Total	180	95	52.77%

Table 4. The prevalence rate with intestinal parasites according to the gender of the goat.

Mix infection	No. of Samples Examined	No. of positive	prevalence (%)
<i>Giardia sp</i> + <i>Cryptosporidium sp</i>	180	3	1.66%
<i>Toxocara vitulorum</i> + <i>Giardia sp</i>		3	1.11%
<i>Toxocara vitulorum</i> + <i>Cryptosporidium sp</i>		4	2.22%

Table 5. Prevalence of mixed infection intestinal parasite in goat.



Figure 1. Oocyst of *Cryptosporidium* sp.



Figure 2. Cyst of *Amoeba* sp.

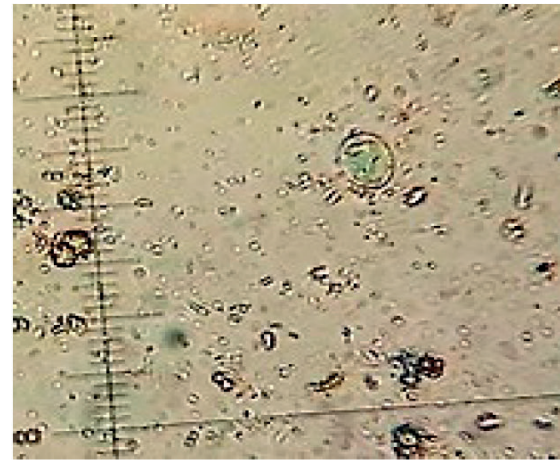


Figure 3. Cyst of *Giardia* sp.



Figure 4. Egg of *Toxocara vitulorum*.

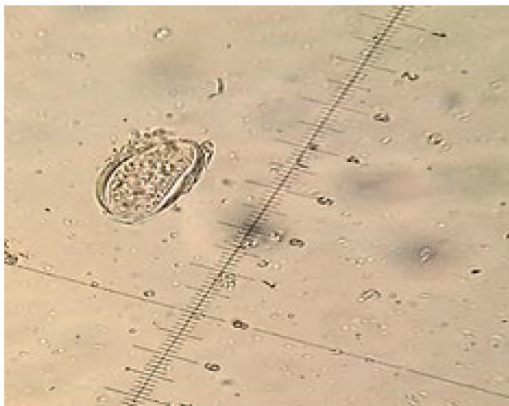


Figure 5. Egg of *Paramphistomum cervi*.

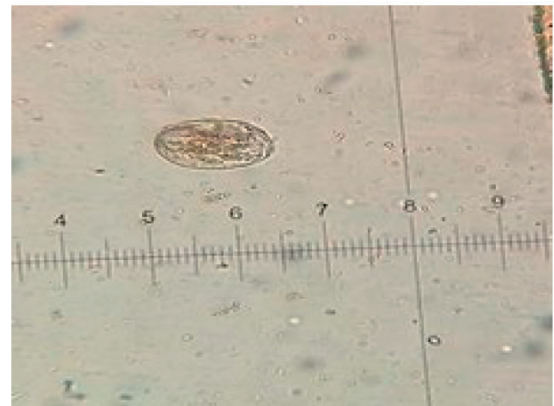


Figure 6. Egg of *Distoma dendriticum*.

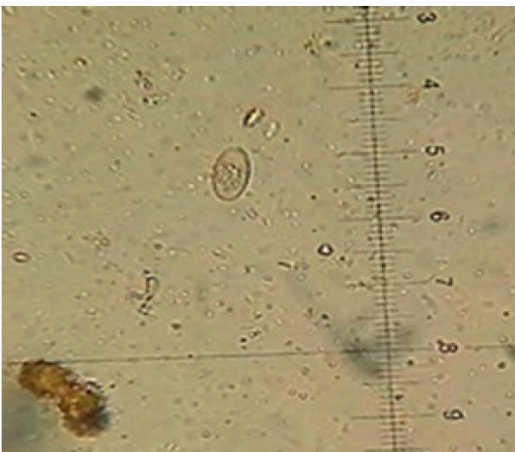


Figure 7. Egg of *Trichostrongylus* sp.

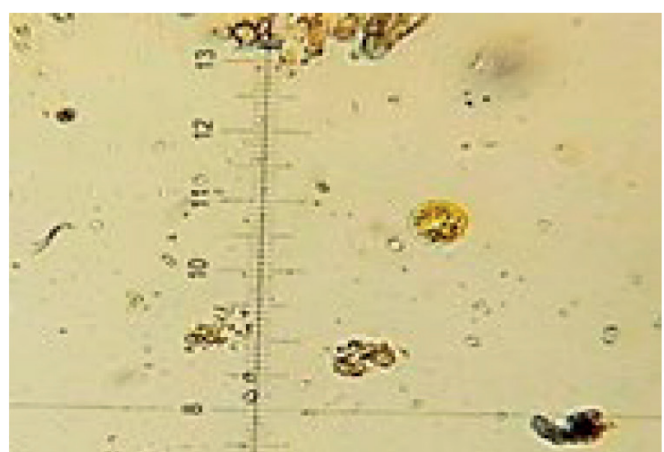


Figure 8. Oocysts of *Cyclospora* sp.



Figure 9. Cyst of *Balantidium coli*.



Figure 11. *Filariae*.

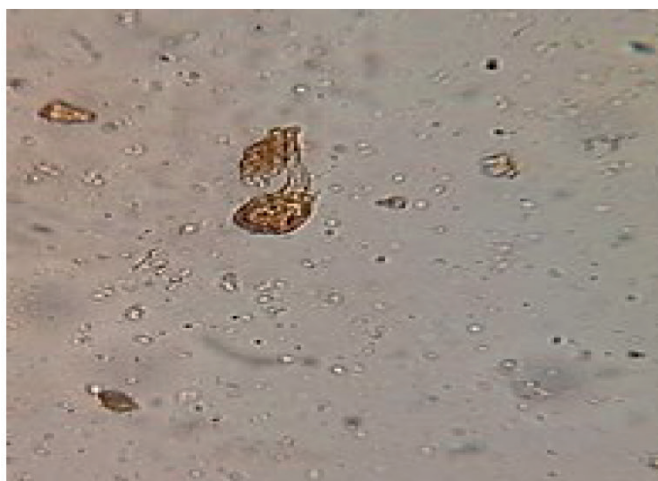


Figure 10. *Cercaria* larvae.

managed in Conventional systems in which large numbers of animals are routinely kept together. This could be due to increased pasture contamination or to poor sanitation and reduced immunity. This may increase the prevalence of intestinal parasites among the animals.

Conclusions

The current study identified several species of intestinal parasites found in goats in Wasit province. This is a significant issue for livestock. As a result, a serious strategy is needed to prevent the spread of more intestinal parasites among local goats and other animals (sheep & cattle) because many parasites may be transmitted between these animals. Furthermore, studies on intestinal parasites in various parts of the country are needed to assess their importance as a source of health hazards. To reduce the parasite burden, some control measures for gastrointestinal parasites in small ruminants must be implemented. Grazing fields should be kept clean and free of animal feces and urine. Education of goat owners on the process of transmission and the influence of these parasites on farm animal productivity should be done regularly.

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Conflicts of Interest

The authors declare no conflict of interest.

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According to the data, the prevalence rate of infection with an intestinal parasite is high. It included: *Toxocara vitulorum* (16.66%), *Cryptosporidium spp* (11.11%), *Amoeba sp* (8.8%), *Giardia sp* (8.8%), *Trichostrongylus sp* (8.33%), *Cyclospora sp* (5.55%), *Dicrocoelium dendriticum* (*Distoma dendriticum*) (5%), *Paramphistomum cervi* (4.44%), cercaria larva (2.22%), *Balantidium coli* (1.66%), *Filariae* form larvae (1.66%), respectively. This study agreed with (14), who found the highest prevalence of Gastrointestinal parasites in goats (91.55 %) in Kirkuk province, Iraq. And (15) found that infection of goats with gastrointestinal parasites was (81.81 %) in Diyala Province, Iraq. In the current study, the lowest infection rate was observed in December (33.33 %), while the highest was in April (70 %). This study is similar to others^{8,16,17}, which found that the wet months had a higher incidence of infection than the winter months. Maybe a related reason is that reduced grazing hours also reduce the chances of contact between the host and parasites, leading to lower prevalence in the winter season. Further, inclement environmental conditions in winter resulting reduced egg production. According to sex this study recorded infection in females more than in males, agreed with (18), who attributed this variation to a physiological status like pregnancy/ lactation, which causes a dip in natural body resistance in goats against parasites. While (19) and (16) also reported a higher prevalence of gastrointestinal parasites in females than in males. (20) and (21), on the other hand, demonstrated that animal sex did not affect the prevalence of gastrointestinal parasites in small ruminants. In contrast to the current findings, (22) found that males had a higher prevalence and intensity of infection than females. The presence of mixed infection with the gastrointestinal parasites in this study has previously been reported in these animal species^{13-15,23}. Perhaps the reason is that goats are

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